# DEHYDRODIEUGENOLS FROM OCOTEA CYMBARUM\*

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**Abstract**—The trunk wood of *Ocotea cymbarum* from the Amazon basin contains  $\alpha$ -phellandrene,  $\alpha$ -pinene, eugenol, dehydrodieugenol and its monomethyl ether, as well as the previously unknown dehydrodieugenol-B (4,5'-diallyl-2'-hydroxy-2,3'-dimethoxydiphenyl ether).

#### INTRODUCTION

During World War II, subsequent to the interruption of trade with Japan, 'oil of sassafras' from Santa Catarina State, Brazil, became a major source of safrol. Since 'oil of sassafras' refers to the essential oil of Sassafras albidum (Nutt.) Nees (Lauraceae), U.S. Customs officials suggested adoption of a name referring to botanical source and the Brazilian product was exported for many years under the designation 'essential oil of Ocotea cymbarum' [2]. The correct binomial for the above species is actually O. pretiosa (Nees) Mez (Lauraceae), but at that time this was rejected by botanists, due to the fact that Mez had stressed the strong cinnamon odour of its wood as a predominant character [3]. Sassafras wood from Santa Catarina, however, smells strongly of safrol. The phenomenon was later shown to be due to physiological variation and chemical aspects were elucidated [4-6].

We have now been able to collect a sample of authentic *Ocotea cymbarum* (H.B.K.) Nees and here report the analysis of its chemical constituents.

## RESULTS

O. cymbarum occurs in the Amazon basin. Vapour entrainment of its trunk wood gave an essential oil consisting chiefly of  $\alpha$ -phellandrene, accompanied by its decomposition product p-cymene, as well as by  $\alpha$ -pinene and by trace amounts of  $\beta$ -pinene and eugenol. Safrol was not detected.

Solvent extraction of the wood led to three compounds, dehydrodieugenol (1a), previously isolated from Litsea turfosa, an Indian Lauracea [7], its monomethyl ether (1b) and a novel oxidative dimer of eugenol designated dehydrodieugenol-B (2a). Dehydrodieugenol (1a) was identified by direct comparison with a synthetic sample [8, 9].

1a  $R^1 = R^2 = H$ 

**1b**  $R^1 = H$ ,  $R^2 = Me$ 

1c  $R^1 = Ac$ ,  $R^2 = Me$ 

**1d**  $R^1 = R^2 = Me$ 

The novel O-methyldehydrodieugenol (1b) gave an acetate (1c) which helped structural characterization by spectral means. As expected, the synthetic mono-O-methyl derivative of 1a was identical with the natural compound (1b). The MS of natural 1b contained a relatively feeble peak at m/e 354. The dimethyl ether of 1a, di-O-methyldehydrodieugenol (1d), may thus also occur in nature.

Dehydrodieugenol-B (2a) was recognized as an isomer of 1a by high resolution MS. The oxygen and allyl substitution of the aromatic rings was assigned by 270 MHz <sup>1</sup>H NMR and confirmed by double irradiation. An allylmethoxybenzene was identified as the major fragment by high resolution MS. Its formation is best explained by a 1,6-hydrogen rearrangement, which requires a free hydroxyl to be situated vicinal to the bridge position of the trioxygenated ring. Only two structures, 2a and 3a, are compatible with these results. Acetylation of the hydroxyl in 3a would be

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expected to shift at least the *ortho*-hydrogen signal about  $0.3 \, \text{ppm}$  to lower field. As the observed shifts are relatively slight  $(-0.02 \, \text{and} \, -0.12 \, \text{ppm})$ , however, both hydrogens probably keep a *meta*-relation with the hydroxyl, and **2a** is the correct representation.

The mesomeric oxidized eugenol radical  $4 \leftrightarrow 5$  would account for the biosynthesis of both dehydrodieugenol  $(5+5\rightarrow 1a)$  and dehydrodieugenol-B  $(4+5\rightarrow 2a)$ .

#### **EXPERIMENTAL**

Isolation of the constituents. A specimen from a small island of the lower Rio Negro, Amazonas, was collected and identified by Prof. K. Kubitzki, Hamburg. Voucher: Herbarium INPA, Manaus, 58576. A wood sample gave 0.8% of essential oil. Another sample (722 g) was percolated with EtOH giving an extract (46 g). The CHCl<sub>3</sub>-soluble part (25 g) was chromatographed on silica (400 g). Petrol, C<sub>6</sub>H<sub>6</sub> and EtOH in the following proportions 1:0:0, 9:1:0 to 7:3:0, 1:1:0, 0:1:0, 0:98:2 eluted, respectively, terpenes (2.3 g), eugenol (0.7 g), sitosterol (0.3 g) and a mixture (10 g), aliphatic ester (40 mg) and a glycoside (43 mg). The latter two fractions were identical to analogous fractions from Endlicheria anomala Nees (Lauraceae) [10] and were not further characterized. The mixture was crystallized from EtOH to 1a (5.3 g). The mother liquor was evapd and the residue chromatographed on alumina. Petrol eluted 1b (2.8 g) and an additional product, which upon prep. TLC (Al<sub>2</sub>O<sub>3</sub>, petrol) gave 1b, 2a (25 mg) and aliphatic oil (30 mg).

Dehydrodieugenol (1a). Mp and lit. [7] mp 106-107°. Identified by direct comparison with a synthetic sample. Methyl ether (1b), 1a (120 mg), Me<sub>2</sub>SO<sub>4</sub> (3 ml), dry K<sub>2</sub>CO<sub>3</sub> (250 mg) and dry Me<sub>2</sub>CO (30 ml) were heated under reflux (5 hr). Work-up gave 1b (100%), identical with the natural product below. Detectable <sup>1</sup>H NMR amounts of dimethyl ether were not produced, even upon extension of the reflux time to 10 hr.

*O-Methyldehydrodieugenol* (**1b**). Oil,  $\lambda_{\text{max}}^{\text{MeOH}}$  nm: 270 inf., 283 (ε 4200, 6500);  $\nu_{\text{max}}^{\text{film}}$  cm<sup>-1</sup>: 3450, 1642, 1592, 1497, 1471, 1429, 1282, 1250, 1159, 1070, 935. <sup>1</sup>H NMR (60 MHz, CDCl<sub>3</sub>): δ 3.33 (d, J = 7 Hz, 2CH<sub>2</sub>), 3.60 (s, OMe), 3.86 (s, 2 OME), 4.8–5.3 (m, 2 CH<sub>2</sub>), 5.7–6.4 (m, 2 CH), 6.63 and 6.70 (2 s, 4 ArH). MS (m/e): 340 (100%) M<sup>+</sup>, 178 (4), 177 (3), 164 (8), 163 (5). Acetate (**1c**). Oil,  $\nu_{\text{max}}^{\text{him}}$  cm<sup>-1</sup>: 1767, 1639, 1587, 1471, 1429, 1294, 1220, 1163, 1075, 1031, 939, 870. <sup>1</sup>H NMR (60 MHz, CCl<sub>4</sub>): δ 2.0 (s, OAc), 3.4 (d, J = 7 Hz, 2 CH<sub>2</sub>), 3.5 (s, OMe), 3.83 (s, 2 OMe), 4.9–5.3 (m, 2 CH<sub>2</sub>), 5.5–6.3 (m, 2 CH). 6.59 and 6.66 (2 d, J = 2 Hz, 2 ArH), 6.72 (s, 2 ArH).

Dehydrodieugenol-B (2a). Oil,  $\lambda_{\text{max}}^{\text{McOh}}$  nm: 275 infl., 285, 325 infl. ( $\varepsilon$  9050, 9200, 5000);  $\nu_{\text{mas}}^{\text{film}} \text{ cm}^{-1}$ : 3460, 1667, 1592, 1508, 1456, 1422, 1267, 1212, 1134, 1091, 1038, 998, 921. <sup>1</sup>H NMR (60 MHz,  $CCl_4 + CDCl_3$ ):  $\delta$  3.21 and 3.33 (2 d, J = 7 Hz, 2 CH<sub>2</sub>), 3.81 and 3.85 (2 s, 2 OMe), 4.9–5.2 (m, 2  $CH_2$ ), 5.7–6.2 (m, 2 CH), 6.25 and 6.38 (d, J = 2 Hz, 2 ArH), 6.71 (s, 3 ArH). MS (m/e): 326.1520 (100%,  $C_{20}H_{22}O_4$ requires 326.1518) M<sup>+</sup>, 177 (8), 167 (60), 164 (7), 163 (9), 149 (19), 148.0809 (60%, C<sub>10</sub>H<sub>12</sub>O requires 148.0888), 147 (17), 133 (8), 131 (10), 117 (14), Acetate (2b), Oil,  $\nu_{\text{max}}^{\text{film}} \text{ cm}^{-1}$ : 1773, 1684, 1605, 1511, 1462, 1434, 1364, 1269. 1193, 1100, 1042, 1011, 918. <sup>1</sup>H NMR (270 MHz, CDCl<sub>3</sub>):  $\delta$  2.24 (s, OAc), 3.25 and 3.37 (2 d, J = 6.5 Hz, 2 CH<sub>2</sub>), 3.81 and 3.82 (2 s, 2 OMe), 5.0-5.1 (m, 2 CH<sub>2</sub>), 5.8-6.1 (m, 2 CH), 6.27 and 6.50 (2 d, J = 1 Hz, 2 ArH), 6.70 (dd, J = 1.5, 8 Hz, ArH), 6.79 (d, J = 1.5 Hz, ArH), 6.87 (d, J = 8 Hz, ArH)

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### REFERENCES

- Ferreira, Z. S., Gottlieb, O. R. and Roque, N. F. (1980) Biochem. Syst. Ecol. 8, 51.
- Guenther, E. (1950) The Essential Oils, Vol. IV, p. 199. Van Nostrand, New York.
- Mez, C. (1889) Jahrbuch des Koeniglichen botanischen Gartens und des botanischen Museums zu Berlin, Vol. 5. p. 250. Gebrueder Borntraeger & Eggers, Berlin.
- Mors, W. B., Magalhães, M. T. and Gottlieb, O. R. (1959) Perf. Essent. Oil Rec. 50, 26.
- Gottlieb, O. R. and Magalhães. M. T. (1960) Perf. Essent. Oil Rec. 51, 18.
- Gottlieb, O. R., Fineberg, M. and Magalhães, M. T. (1962) Perf. Essent. Oil Rec. 53, 219, 299.
- 7. Holloway, D. M. and Scheinmann, F. (1973) Phytochemistry 12, 1503.
- Cousin, H. and Hérissey, H. (1908) Bull. Soc. Chim. Fr., Ser. 4 3, 1066.
- 9. Fujita, Y. and Shigenoi, J. (1966) Nippon Kagaku Zasshi 87, 1002: apud Chem. Abstr. 65, 18450.
- Diaz, A. M. P. de, Diaz, D., P. P., Ferreira, Z. S., Gottlieb, O. R., Lima, R. A. de and Cavalcante, S. de H. (1977) Acta Amazonica (Manaus) 7, 292.